

Multispecies probiotics applications through feed addition on cultured tilapia to prevent streptococcosis

Aplikasi probiotik multispecies melalui pakan pada budidaya ikan nila *Oreochromis niloticus* untuk pencegahan streptococcosis

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ABSTRACT

Streptococcosis is one of the diseases that often arise and can cause death by 100% in tilapia fish farming. This study aimed to determine the most optimal probiotics combination of cells *Bacillus cereus* P22, *Bacillus subtilis* ND2 and *Staphylococcus lentus* L1k through the feed for streptococcosis prevention. The experimental fish with initial body weight of (13.06±0.18 g), were randomly distributed at 25 fish per 60 L water, and reared for 21 days. The fish fed with supplemented feed (except controls) with FR 3%, about three times a day, for 14 days. The treatment of combination probiotics-supplemented feed on, K1 (P22 and ND2, density of 10⁶ CFU/mL), K2 (ND2 and L1k, density of 10⁵ CFU/mL), K3 (P22 and L1k, density of 10⁵ CFU/mL), K4 (P22, ND2 and L1k, density of 10⁶ CFU/mL), the negative and positive control treatment given feed without supplementation. On day 15, the experimental fish, (except negative control) were challenged with *Streptococcus agalactiae* via intraperitoneal injection 0.1 mL/fish (10⁴ CFU/mL). The results indicated that the best combination was shown by K3 (the combination of *Bacillus cereus* P22 and *Staphylococcus lentus* L1k) with cells addition 1% (v/v). The result of K3 survival rate shown 89.33±6.11%, significantly different from the K+ and K- which were 66.67±4.62% and 88.00±8.00%. The highest increased immune value shown by K3 were which phagocytic index value 64.00±2.65% and respiratory burst activity value 0.07±0.00 OD.

Keywords: multispecies probiotics, *Oreochormis niloticus*, streptococcosis

ABSTRAK

Streptococcosis merupakan salah satu penyakit yang seringkali muncul dan dapat menyebabkan kematian hingga 100% pada budidaya ikan nila. Penelitian ini bertujuan untuk menentukan kombinasi optimal dari probiotik *Bacillus cereus* P22, *Bacillus subtilis* ND2, dan *Staphylococcus lentus* L1k melalui pakan sebagai upaya pencegahan streptococcosis. Ikan uji berukuran (13,06±0,18 g), secara acak dipelihara dengan kepadatan 25 ekor/ akuarium dengan volume air 60 L, dipelihara selama 21 hari dan diberi pakan yang disuplementasi probiotik (kecuali kontrol) dengan perkiraan FR 3% sebanyak tiga kali sehari, selama 14 hari. Perlakuan kombinasi probiotik yang disuplementasi pada pakan yaitu: K1 (P22 dan ND2, kepadatan 10⁶ CFU/mL), K2 (ND2 dan L1k, kepadatan 10⁵ CFU/mL), K3 (P22 dan L1k, kepadatan 10⁵ CFU/mL), K4 (P22, ND2 dan L1k, kepadatan 10⁶ CFU/mL), perlakuan kontrol negatif dan positif diberi pakan tanpa suplementasi probiotik. Pada hari ke-15, ikan uji (kecuali kontrol negatif) diinjeksi dengan *Streptococcus agalactiae* dengan kepadatan 10⁴ CFU/mL sebanyak 0,1 mL secara intraperitoneal. Hasil penelitian ini menunjukkan bahwa kombinasi probiotik multispecies terbaik adalah perlakuan K3 (kombinasi *Bacillus cereus* P22 dan *Staphylococcus lentus* L1k) dengan dosis 1% (v/v). Nilai kelangsungan hidup K3 didapat 89,33±6,11%, berbeda nyata dengan K+ (66,67±4,62%) dan K- (88,00±8,00%). Nilai respons imun dengan peningkatan tertinggi pada perlakuan K3 dengan nilai indeks fagositik 64,00±2,65% dan aktivitas respiratory burst 0,07±0,00 OD.

Kata kunci: probiotik multispecies, *Oreochormis niloticus*, streptococcosis

INTRODUCTION

Streptococcosis is a disease that often arises in Tilapia farming activities and is caused by a pathogen called *Streptococcus agalactiae* (Tauhid *et al.*, 2014). *S. agalactiae* pathogen attacks are acute in tilapia (Hernandez *et al.*, 2009). The outbreaks of *S. agalactiae* infection were first noticed in Malaysian (Nijah *et al.* 2012), Asian and Pacific Asian waters. In 2008, streptococcosis was reported to cause tremendous economic losses, which reached approximately \$250 million (Klesius *et al.*, 2008). In addition, a case of streptococcosis infection in Karawang (West Java) resulted in mortality rates of up to 50% on tilapia farms (Gardenia *et al.*, 2011). Visual clinical symptoms of *S. agalactiae* infection included detaching scale and abnormal swimming motions (Ye *et al.*, 2011), bulging eyes (exophthalmos) (Filho *et al.*, 2009), bulged stomach and bleeding on the body. Changes in the anatomy of internal organs that can occur are liver dysfunction, swelling spleen, and brain dysfunction (Hardi *et al.*, 2011; Li *et al.*, 2014).

Probiotic is defined as living microorganisms that once provided in sufficient amount, can positively affect the microbiota balance of the intestine, leading to increment in the functions of the host immune system (Sanders, 2008). The application of probiotics is an attempt in preventing the infection of cultured fish by disease (Flores, 2011). Widyaningsih (2011), stated that probiotics positively influence the immune system. Two main techniques have been used in the application of probiotics i.e. through the rearing medium or the feed. Several types of research have proved that probiotic application can benefit tilapia farming by increasing immune system and resistance against diseases (Aly *et al.*, 2008a), so as to increase the survival rate of the fish (Aly *et al.*, 2008b). According to Sya'bani *et al.* (2015), the supplementation of *Bacillus* sp. and *Staphylococcus lentus* (every 2 days) at a dose of 10^3 CFU/mL through the rearing medium of catfish can result in a survival rate of 93.33%. Meanwhile, results of *Bacillus* sp. probiotic application, at a dose of 10^6 CFU/mL, through the rearing medium could maintain a survival rate of 100% in common carp (Haditomo *et al.*, 2016). According to Fidyandini *et al.* (2016), application of multispecies *Bacillus subtilis* and *Staphylococcus lentus* probiotic through rearing medium of catfish (Dumbo strain) can not only push *Aeromonas hydrophila* cell population up

to 40% compared to the positive control, but also increase survival rate, growth, immune response, and decrease feed conversion ratio as well.

In accordance with the descriptions above, a solution that has been applied to the problem of *S. agalactiae* pathogen infection on the potential of optimal intensive production of tilapia is probiotics. The present research was carried out to determine the best combination of multispecies probiotics that can be supplemented through the feed to prevent streptococcosis.

MATERIALS AND METHODS

Test fish

The test fish used in the present study was BEST (Bogor Enhanced Strain tilapia) tilapia *Oreochromis niloticus*. The fish had an average body length of 9.10 ± 0.08 cm with an average body weight of 13.06 ± 0.18 g and were brought from Plasma Nutfah Research Installation, the center for freshwater research and development, Cijeruk, West Java. Fish were acclimatized in fiber tanks that were equipped with an aeration system and fed three times a day at satiation.

Pathogenic and probiotic bacteria preparation

The bacterium used in the present study was *Streptococcus agalactiae* N14G pathogen, while probiotic bacteria used included *Bacillus cereus* P22, *Bacillus subtilis* ND2, and *Staphylococcus lentus* L1k. N14G, P22, and ND2 isolates were part of collections from the center for research and development of freshwater aquaculture. L1k isolate was a collection from fish health laboratory, at the Faculty of Fisheries and Marine Science, Bogor Agricultural University.

Antibiotic resistance marker was used on isolates that were to be used in the present study in order to determine the presence of the bacterium in the rearing environment, which presence could be monitored with total plate count (TPC) method. Antibiotic used in the present study was Rifampicin (Rif) as *S. agalactiae* N14G marker. *S. agalactiae* N14G inoculant was cultured in 10 mL BHIB (brain heart infusion broth), incubated for 48 hours at a temperature of 28 °C, re-cultured in 10 mL BHIB containing 100 µg/mL of Rifampicin (Rif), and finally incubated for 48 hours. The N14G Rif^R that grew was inoculated on BHIA (brain heart infusion agar) containing the same dose of antibiotics.

An antibiotic that was used as resistance marker for probiotic bacteria was based on Fidyandini

et al. (2016) research i.e. ciprofloxacin (Cip) as P22 marker, cefadroxil (Cef) as ND2 marker, tetrasiklin (Tet) as L1k marker with an antibiotic dose of 100 µg/mL. Antibiotic resistance marking process was carried out by culturing each isolate on TSB (tryptic soy broth) medium and incubating for 24 hours at a temperature of 28 °C. Each grown isolate was re-cultured on TSB containing 100 µg/mL of antibiotic and incubated for 24 hours at 28 °C. Resistant isolates that have grown were re-cultured each on TSA (tryptic soy agar) medium containing the same dose of antibiotic and measurements of overflow in pathogenic and probiotic isolates were carried out through spread method (Madigan *et al.*, 2011).

Test feed preparation

Commercial feed (FF 999) with a protein content of 38% was used in the present study. The prepared probiotics combinations that were supplemented in the feed were as follows: K1 (probiotic combination of P22 Cip^R 10⁶ CFU/mL and ND2 Cef^R 10⁶ CFU/mL), K2 (probiotic combination of ND2 Cef^R 10⁵ CFU/mL and L1k Tet^R 10⁵ CFU/mL), K3 (probiotic combination of P22 Cip^R 10⁵ CFU/mL and L1k Tet^R 10⁵ CFU/mL), K4 (probiotic combination of P22 Cip^R 10⁶ CFU/mL, ND2 Cef^R 10⁶ CFU/mL and L1k Tet^R 10⁶ CFU/mL), meanwhile K+ (positive control) and K (negative control) were not supplemented with probiotic combinations. The process of pellet formulation included mixing multispecies probiotics, at a dose of 1% ($\frac{1}{b}$) (Setiawati *et al.*, 2013), and white egg, 2% ($\frac{2}{b}$), which was added with a 6% ($\frac{6}{b}$) distilled water. Afterward, the solution was homogenized, evenly mixed with the feed, and dried.

Fish rearing

Rearing facilities used in the present study were 24 aquariums (65×35×35 cm³) with a capacity of 60 Liters of water each. The stocking density of test fish was 25 fish/aquarium. Fish were fed on a 38% protein feed and daily feeding rate (FR) was maintained at 3% of the test fish biomass. Feeding was performed three times a day while feeding with probiotic supplemented feed was performed once a day (in the morning) for 14 days.

Water quality in the rearing facility was maintained at an optimum level by mean of discharging and water exchange (50%) every two days. Water quality parameters such as dissolved oxygen, pH, and ammonia were measured throughout the research. Meanwhile, water

temperature was daily measured. The ranges of water quality parameters during the present research were as follows: DO (3.9–5.2 mg/L), pH (6.3–7.5) and ammonia (0.001–0.0147 mg/L), while water temperature ranged between 27 and 29 °C. The ranges of water quality parameters in the present study were in line with the standard ranges for tilapia culture as described by the national standards (SNI 7550: 2009 (2009).

Challenge test

Tilapia that were given treatments for 14 days were injected (intraperitoneal) with *S. agalactiae* pathogenic bacteria on day 15 (Positive control, K1, K2, K3, and K4) with at a concentration of 10⁴ CFU/mL and a dose of 0.1 mL/fish. Fish in the negative treatment were injected with PBS at 0.1 mL/fish and mortality was observed over a period of 7 days.

Test parameters

The test parameters were as follows: growth performances such survival rate (SR) (Effendi, 2002), specific growth rate (SGR) (Huisman, 1987), feed conversion ratio (FCR) (Takeuchi, 1988), that were measured throughout the research. The immune response was observed by mean of collecting blood samples from 3 fish in each treatment. Blood samples were collected on day 0, 7, 14 (pre-challenge test), day 17, 19 (post challenge test), until the end of the research period on day 21. The immune response involved parameters such as phagocytic activity (PA) (Anderson & siwicki, 1995), and respiratory burst (RB) based on Secombes (1990) method that was modified according to Stasiack and Baumann (1996) in Singh *et al.* (2013). Monitoring of pathogenic cell population was performed on day 17, 19, and 21. Monitoring of probiotic cell population was performed on day 3, 6, 9, 12, 15, 18, and 21. Monitoring of both pathogenic and probiotic cell population was performed through Cawan sebar method (Madigan *et al.*, 2012).

Data analysis

A completely randomized design with 6 treatments was used in this study and each treatment was replicated 4 times. Data on SR, SGR, FCR, PA, RB, were analyzed using ANOVA test (on SPSS ver. 20) at an interval of P<0.05. An ANOVA test was followed by Duncan's post-hoc comparison test if significant differences were found. Monitoring of pathogenic and probiotic bacteria abundance was descriptively analyzed.

RESULTS AND DISCUSSION

Results

Survival rate and growth performance

K3 treatment had the highest survival rate ($89.33 \pm 6.11\%$) which was significantly different ($P < 0.05$) compared to treatment K+ ($66.67 \pm 4.62\%$). In addition, K3 treatment had the highest SGR ($2.37 \pm 0.02\%$) as well that significantly differed to other treatments (including control). The lowest FCR was observed in K3 treatment ($1.26 \pm 0.03\%$), but was not significantly different to K1 treatment ($1.30 \pm 0.03\%$). However, the FCR of the two mentioned treatments were significantly different ($P < 0.05$) to those of control, K2 ($1.32 \pm 0.03\%$), K4 ($1.36 \pm 0.04\%$), which were in turn significantly different to control (Table 1).

Phagocytic activity (PA)

The observation of the phagocytic activity (PA) in the test fish blood (in each sampling) resulted in fluctuating outcomes. The highest PA activity was observed in treatment K3 during the rearing period and significantly differed to ($P < 0.05$, Figure 1) both other treatments and control.

Respiratory burst activity (RB)

The respiratory burst activity after 14 days of probiotics supplementation resulted in significant differences among treatments and control ($P < 0.05$). The highest respiratory burst was shown in treatment K3 which was significantly different to other treatment and control ($P < 0.05$, Figure 2) during the 21 days rearing period.

Monitoring of pathogenic and probiotic cells population

S. agalactiae N14G Rif^R observation results on the brain (Figure 3A), the eyes (Figure 3B) and the kidney (Figure 3C) of the test fish showed that the fish in each treatment, that was supplemented with

multispecies probiotics, was able to significantly press the growth of *S. agalactiae* N14G Rif^R ($P < 0.05$) compared to control. Treatment K3 had the best results in terms of multispecies probiotic combination (P22 Cip^R and L1k Tet^R).

The results of the total probiotic cells observation during the rearing period in the intestine of the test fish (Figure 4A) showed fluctuation between probiotic treatments (K1, K2, K3, and K4), however, all the probiotic treatments experienced significant increments.

Discussion

The results of the present study on tilapia fed on treatments K1, K2, K3, and K4 showed positive responses, leading to survival rates that were not significantly different among the four probiotic treatments. However, the survival of the probiotic treatments were significantly different ($P < 0.05$; Table 1) compared to that of treatment K+, indicating that supplementation of probiotics through feed can increase the immune response, leading to an increase in survival. Nevertheless, the highest survival was observed in treatment K3 (89.33%). It was believed that the K3 probiotic combination treatment (P22 Cip^R and L1k Tet^R), with a dose of 1% and a concentration of 10^5 CFU/mL, was able not only of expanding the immune response, but also providing a better pressing against the growth of *S. agalactiae* N14G Rif^R pathogen. Indeed, the results of monitoring the abundance of pathogens in targeted organs of the test fish with *S. agalactiae* N14G Rif^R pathogenic bacteria demonstrated that treatment K3 had the lowest pathogen abundance (Figure 3). According to Haditomo *et al.* (2016), the supplementation of *Bacillus* sp. probiotic through the rearing medium can maintain the survival of common carp at 100%. In addition, Wang *et al.* (2016) stated that supplementing *Bacillus cereus* through both medium and feed can improve the health status of the fish. However, supplementation through feed

Table 1. Survival rate (SR), specific growth rate (SGR) and feed conversion ratio (FCR)

Parameters	Treatments					
	K-	K+	K1	K2	K3	K4
SR (%)	88.00±8.00b	66.67±4.62a	86.67±6.11b	88.00±8.00b	89.33±6.11b	85.33±8.33b
SGR (%/day)	2.04±0.04b	1.85±0.04a	2.31±0.03d	2.24±0.02c	2.37±0.02e	2.22±0.02c
FCR (%)	1.45±0.05d	1.59±0.02e	1.30±0.03ab	1.32±0.03b	1.26±0.03a	1.36±0.04cb

Note: Negative control (K-), Positive control (K+), P22 Cip^R and ND2 Cef^R combination (K1), ND2 Cef^R (K2) and L1k Tet^R combination (K2), P22 Cip^R and L1k Tet^R combination (K3) and P22 Cip^R, ND2 Cef^R and L1k Tet^R combination (K4). Data on the same line with different letters showed significant differences among treatments ($P < 0.05$).

is a more effective way in improving the immune system of tilapia.

The application of probiotic combination in the four treatments, as observed on the growth response of the test fish, showed significant differences in terms of growth rate among probiotic treatments ($P < 0.05$; Table 1) compared to control. Treatment K3 had the highest SGR that was significantly different to ($P < 0.05$; Table 1) to treatments K1, K2, K4, and control. In addition, the best FCR was also observed in treatment K3 (Table 1), indicating that probiotic bacteria combination supplemented in the feed (that was consumed by the fish) was able to reach and adhere to the digestive tract. This was in line with the probiotic population monitoring data

(Figure 4) and helped in supporting the digestive process. Treatment K+ had the lowest SGR, which was influenced by both total feed intake and nutritional status of the fish before and after the challenge test with *S. agalactiae* N14G Rif^R. The low FCR that were observed in P22 Cip^R and L1k Tet^R probiotic combination treatments illustrated that a low feed consumption (a specific amount of feed) can be optimally utilized so as to increase growth. Salamah *et al.* (2013) reported that the addition of L1k heterotrophic bacteria cells can result in a better survival rate on the host. That was in line with Sya'bani *et al.* (2015) who stated that the application of L1k probiotic combination is able to influence the survival rate (higher percentage) of the host. *B. cereus* P22, *S.*

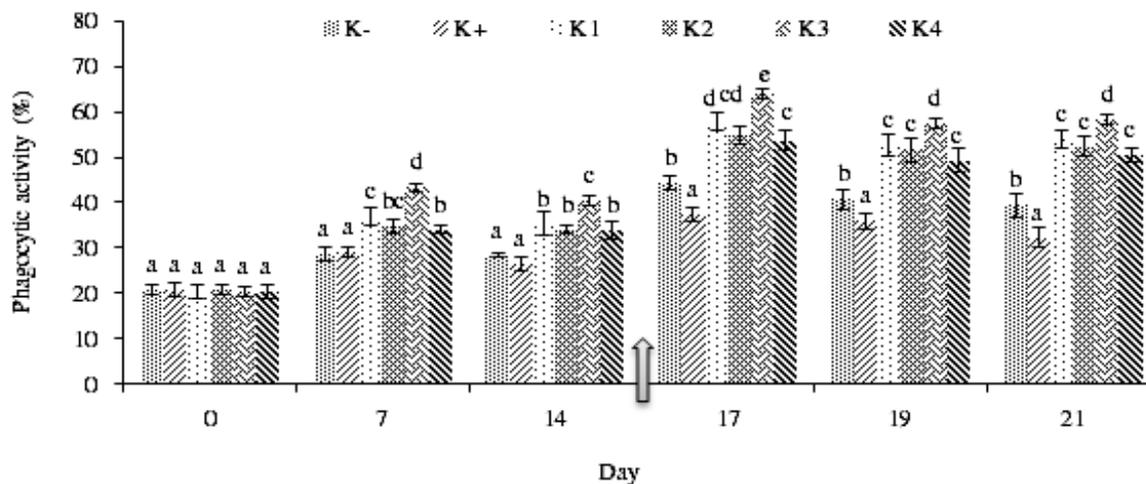


Figure 1. Phagocytic activity (PA) during the rearing period. The arrow shows the injection time of the challenge test with *S. agalactiae* N14G Rif^R. Different letters at the end point of the bar graph showed significant differences ($P < 0.05$). Negative control without challenge test (K-), while positive control (K+), P22 Cip^R and ND2 Cef^R combination (K1), ND2 Cef^R (K2) and L1k Tet^R combination (K2), P22 Cip^R and L1k Tet^R combination (K3) and P22 Cip^R, ND2 Cef^R and L1k Tet^R combination (K4) were subjected to challenge test.

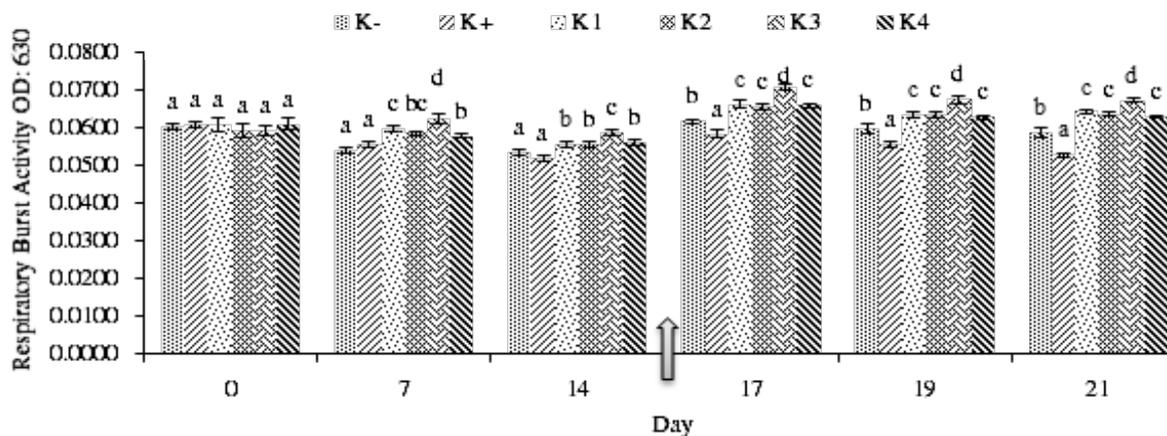


Figure 2. Respiratory burst activity (RB) during the rearing period. The arrow shows the injection time of the challenge test with *S. agalactiae* N14G Rif^R. Different letters at the end point of the bar graph showed significant differences ($P < 0.05$). Negative control without challenge test (K-), while positive control (K+), P22 Cip^R and ND2 Cef^R combination (K1), ND2 Cef^R (K2) and L1k Tet^R combination (K2), P22 Cip^R and L1k Tet^R combination (K3) and P22 Cip^R, ND2 Cef^R and L1k Tet^R combination (K4) were subjected to challenge test.

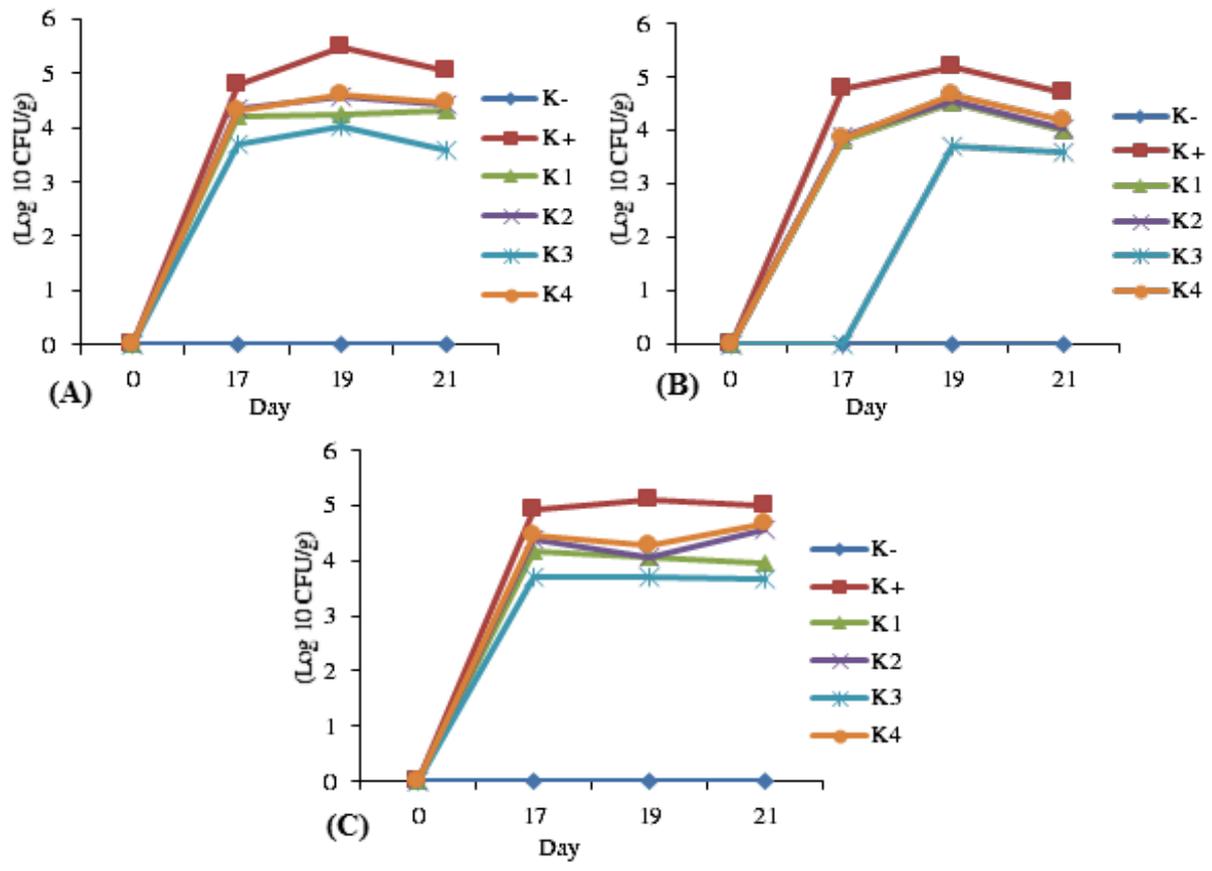


Figure 3. Monitoring of the density of *S. agalactiae* N14G RifR cells in the brain (A), eyes (B), and the kidney (C) (Log 10 CFU/g) of the test fish post challenge test during the rearing period. Negative control without challenge test (K-), while positive control (K+), P22 CipR and ND2 CefR combination (K1), ND2 CefR (K2) and L1k TetR combination (K3), P22 CipR and L1k TetR combination (K4) were subjected to challenge test.

lentus L1k and *B. subtilis* ND2 are Gram-positive bacteria and according to Buruiana *et al.* (2014), Gram-positive bacteria can influence enzymatic activity by increasing it.

Putra and Widanarni (2015) demonstrated that *Bacillus* probiotic supplemented through feed can increase amylase activity, carbohydrate digestion, and growth performance of tilapia. Fidyandini *et al.* (2016), stated that a combination of *S. lentus* L1k and *B. subtilis* ND2 can optimally inhibit the growth of *A. hydrophila* in catfish (*Dumbo* strain). In addition, Lallo *et al.* (2007, 2008), also stated that *B. cereus* can obstruct the growth of *A. hydrophila*. According to Bernard *et al.* (2013), *B. cereus* revealed antagonistic activities against three pathogens i.e. *Vibrio alginolyticus*, *A. hydrophila* and *Pseudomonas putida*, while the strongest antagonistic effect was observed against *V. alginolyticus*. *B. cereus* is safe for usage in tilapia farming activities.

Health status of tilapia was observed based on blood profile parameters as indicators of the immune response. Treatments K1, K2, K3, and

K4 significantly influenced both PA and RB of tilapia compared to treatments K- and K+. PA and RB (Figure 1 and Figure 2), in each treatment during 21 days, experienced fluctuations. The highest PA occurred on day 7 (2 days after challenge test injection), which was observed in treatment K3 i.e. $64.00 \pm 2.65\%$ and this value was significantly different ($P < 0.05$) compared to K+ treatment ($37.33 \pm 1.53\%$). In line with the increase in PA, the respiratory burst experienced sharp increments on day 17 and treatment K3 had the highest RB 0.07 ± 0.00 OD compared to K+ (0.06 ± 0.00 OD). The phagocytic index illustrates the activity of phagocytic cells in carrying out the phagocytosis process (engulfing pathogenic bacteria cells). RB occurs in Phagocytic cells that perform phagocytosis that produces anion (H_2O_2) and super oxidate (OH) that are highly toxic to bacteria, leading to an increase in phagocytic cells ability to destroy pathogenic bacteria (Rawling *et al.*, 2012). The increments in both PA and RB in the present research demonstrated that the supplementation of a combination of

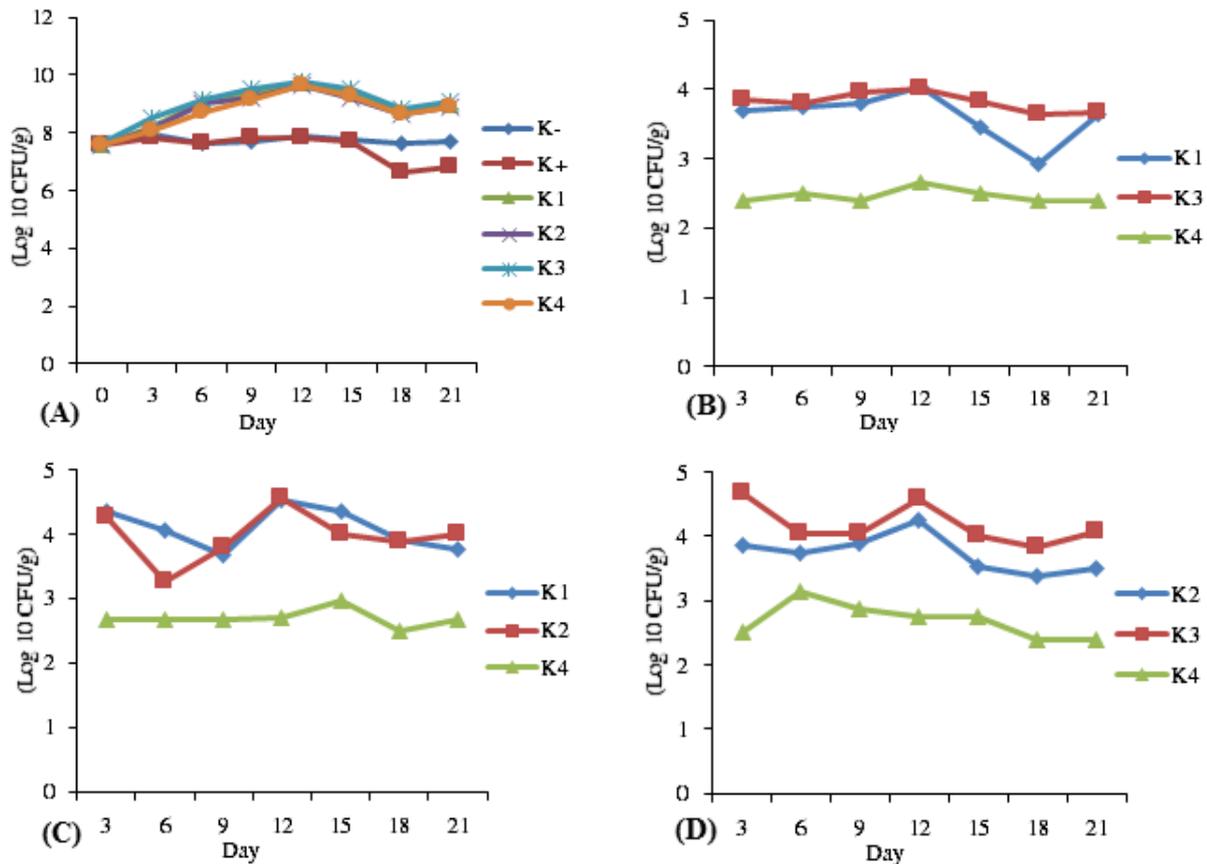


Figure 4. Monitoring of the total probiotic cell density in the intestine (A), the density of P22 CipR (B) cells, density of ND2 CefR probiotic cells (C) and density of L1k TetR (D) probiotic cells (Log 10 CFU/g) of the test fish during the rearing period. Negative control (K-) and positive control (K+) with multispecies probiotics, P22 CipR and ND2 CefR combination (K1), ND2 CefR (K2) and L1k TetR combination (K2), P22 CipR and L1k TetR combination (K3) and P22 CipR, ND2 CefR and L1k TetR combination (K4).

probiotics can enhance the immune system in order to actively stand against *S. agalactiae* N14G Rif^R pathogenic infection. According to Agung *et al.* (2015), changes in blood profile pictures can occur as consequences of both stress condition or presence of pathogens.

The monitoring of cell population displays the ability of the best probiotic combination to inhibit the growth of *S. agalactiae* N14G Rif^R. The lowest cell population density was observed in treatment K3 in the brain (3.56 ± 0.06 Log 10 CFU/g) (Figure 3A), the eyes (3.59 ± 0.03 Log 10 CFU/g) (Figure 3B) and the kidney (3.70 ± 0.17 Log 10 CFU/g) (Figure 3C). In line with that, the observed total probiotic cells experienced increments during 14 days of probiotic combination supplementation (Figure 4A). meanwhile, monitoring probiotic cells P22 Cip^R (Figure 4B), ND2 Cef^R (Figure 4C), and L1k Tet^R (Figure 4D), revealed that the combination of probiotics, that was supplemented in the feed, can adhere to the digestive tract and play a synergic role in supporting increments in immune response, growth response and

maintenance of an optimal survival. The increment and reduction in *S. agalactiae* N14G Rif^R growth are influenced by the population of probiotics. The reduction of pathogen density in the target organs is believed to be the side effect of the increase in probiotics density in the digestive tract of the host.

Nayak (2010) stated that the application of probiotics can increase host resistance against pathogenic infection by mean of increasing the non-specific immune system of the host. Results from several types of research demonstrated that the utilization of heterogeneous probiotics in tilapia and other aquatic organisms can increase the performance of lymphocytes, granulocytes, lysozyme activities and bactericidal serum activity (Pirarat *et al.*, 2011; Sorroza *et al.*, 2012; Sharma *et al.*, 2013). Fidyandini *et al.* (2016) stated that *S. lentus* is a non-pathogenic heterotrophic bacteria so that its supplementation is believed to be able to increase the total bacterial population. *B. cereus* produces anti-microbial substances i.e. cerein (Bizani & Brandelli, 2002) which can

inhibit Gram-positive pathogens (Ghanbari *et al.*, 2009). *Bacillus* spp. makes antibiotic compound i.e colistin and polymixin, that are effective against Gram-negative bacteria (Amin *et al.*, 2012). A research conducted by Navinchandran *et al.* (2014) revealed that extracellular enzymes that are produced by *B. cereus* as probiotics (supplemented through shrimp feed) can increase both immunology parameters and growth of the host. Brito *et al.* (2012) stated that the production of antimicrobial plays a role in the modulation of the immune system i.e. white blood cells so that the performance of spreading white blood cells increases the defense system from pathogenic infections.

CONCLUSION

The appropriate multispecies probiotics combination in the present study was a combination of *Bacillus cereus* P22 and *Staphylococcus lentus* L1k (that was supplemented at the optimal dose of 1 % ($1/w$)) that increased both growth performance and immune response of tilapia to prevent streptococcosis.

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